

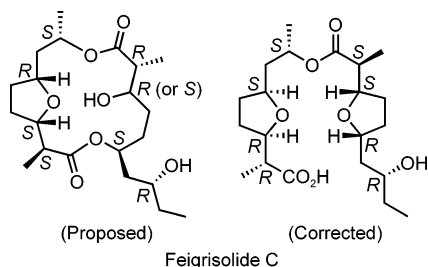
## Feigrisolide C: Structural Revision and Synthesis

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The original macrodiolide structure proposed for feigrisolide C was incorrect. The true structure of feigrisolide C was identified as (2'S,3'S,6'R,8'R)-homononactoyl (2R,3R,6S,8S)-nonactate, which was confirmed by total synthesis.

Feigrisolide C was isolated from the culture broth of *Streptomyces griseus* (strain GT 051022)<sup>1</sup> and was also claimed to be a metabolite of *Streptomyces* sp. 6167 of marine origin.<sup>2</sup> The structure was proposed<sup>1</sup> to be a 15-membered macrodiolide **1** (originally intended to be **2**) incorporating 8-*epi*-nonactate (Figure 1). Both these compounds were synthesized in our laboratory and found to be different from feigrisolide C.<sup>3</sup>

A small sample of natural feigrisolide C was subjected to exhaustive reduction with excess amount of lithium aluminum hydride, and the crude products were acetylated<sup>4</sup> to furnish two products **A** and **B** (Scheme 1).

If one assumes that the true structure of feigrisolide C is epimeric to structure **1** or **2**, then diacetate **3** should represent **A**, and a tetraacetate epimeric to **4** or **5** should correspond to **B**. Surprisingly, examination of the NMR spectra of the major product<sup>5</sup> **B** clearly indicated that it was a diacetate originated from a *threo* homononactate

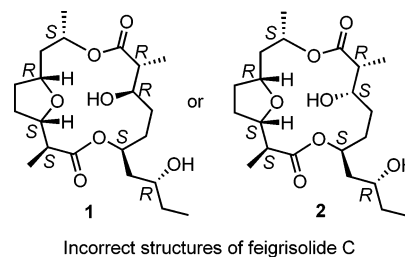
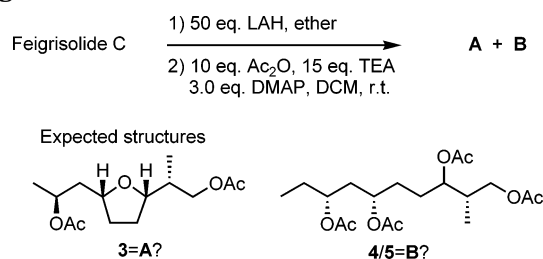


FIGURE 1. Structures proposed for feigrisolide C.

### SCHEME 1. Chemical Transformation of Feigrisolide C



acid with the characteristic ABX signals from C1 methylene group ( $\delta$  3.96 and 4.20,  $\Delta\delta \geq 0.2$ ). Diacetates from *erythro* nonactate acids should exhibit ABX signals different from those of C1 methylene protons;  $\Delta\delta$  0.1.<sup>4</sup>

(+)-Methyl homononactate (**12**) was synthesized from methyl (*R*)-3-hydroxypentanoate (**6**)<sup>6</sup> following the scheme used for previous synthesis of (+)-methyl nonactate (**14**)<sup>7</sup> (Scheme 2). The diacetate **13** obtained from **12** after  $\text{LiBH}_4$  reduction/acetylation exhibited NMR spectra identical to those of **B**.

Examination of the NMR spectra<sup>8</sup> of **A** indicated that it was also a diacetate from a *threo* nonactate. The diacetate **15**, prepared from (+)-methyl nonactate (**14**), furnished NMR spectra identical to those of **A**. For comparison, the alternative diacetate **3** was prepared from (+)-methyl 8-*epi*-nonactate (**16**),<sup>3</sup> and it was clearly different from **A**.

Four candidates for feigrisolide C then emerged: (2'S,3'S,6'R,8'R)-homononactoyl (2S,3S,6R,8R)-nonactate (**17**), *ent*-**17**, (2'S,3'S,6'R,8'R)-homononactoyl (2R,3R,6S,8S)-nonactate (**18**), or *ent*-**18** (Figure 2).

Spectroscopic differences between **17** and **18** were anticipated to be small, and it was decided to prepare both isomers for comparison. The benzyl ether **19** of (2'S,3'S,6'R,8'R)-homononactate acid was prepared from the methyl ester **11**. It was reacted with methyl (2S,3S,6R,8R)-nonactate (**14**) under Yamaguchi conditions to furnish the diester **20**, which was converted into **17** via **21**. Using **19** and *ent*-**14**, we also prepared the other

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(1) Tang, Y.-Q.; Sattler, I.; Thiericke, R.; Grabley, S.; Feng, X.-Z. *J. Antibiot.* **2000**, *53*, 934–943.

(2) Sobolevskaya, M. P.; Fotso, S.; Havash, U.; Denisenko, V. A.; Helmke, E.; Prokofeva, N. G.; Kuznetsova, T. A.; Laatsch, H.; Elyakov, G. B. *Chem. Nat. Compd.* **2004**, *40*, 282–285.

(3) Kim, W. H.; Jung, J. H.; Sung, L. T.; Lim, S. M.; Lee, E. *Org. Lett.* **2005**, *7*, 1085–1087.

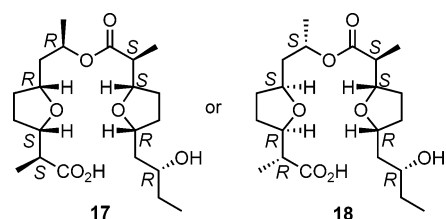
(4) This strategy was used previously in our synthesis of pamamycin-607 to check epimerization problems at C2 and C2' of *seco* acids: Jeong, E. J.; Kang, E. J.; Sung, L. T.; Hong, S. K.; Lee, E. *J. Am. Chem. Soc.* **2002**, *124*, 14655–14662.

(5) The yield of **A** was much lower than that of **B**, and **A** was initially overlooked as TLC separation was difficult.

(6) Noyori, R.; Ohkuma, T.; Kitamura, M.; Takaya, H.; Sayo, N.; Kumobayashi, H.; Akutagawa, S. *J. Am. Chem. Soc.* **1987**, *109*, 5856–5858.

(7) Lee, E.; Choi, S. *J. Org. Lett.* **1999**, *1*, 1127–1128.

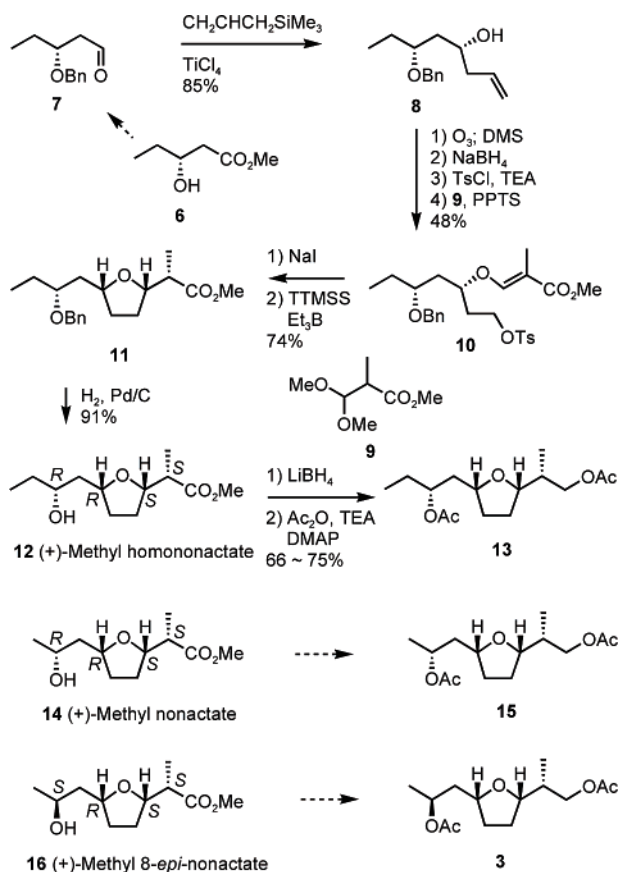
(8) We failed to get clean NMR spectra of **A** due to the lack of material, but the characteristic features were easily recognized.



Possible structures of feigrisolide C  
17, *ent*-17, 18, *ent*-18

FIGURE 2. Possible structures of feigrisolide C.

SCHEME 2. Preparation of Diacetates 3, 13, and 15



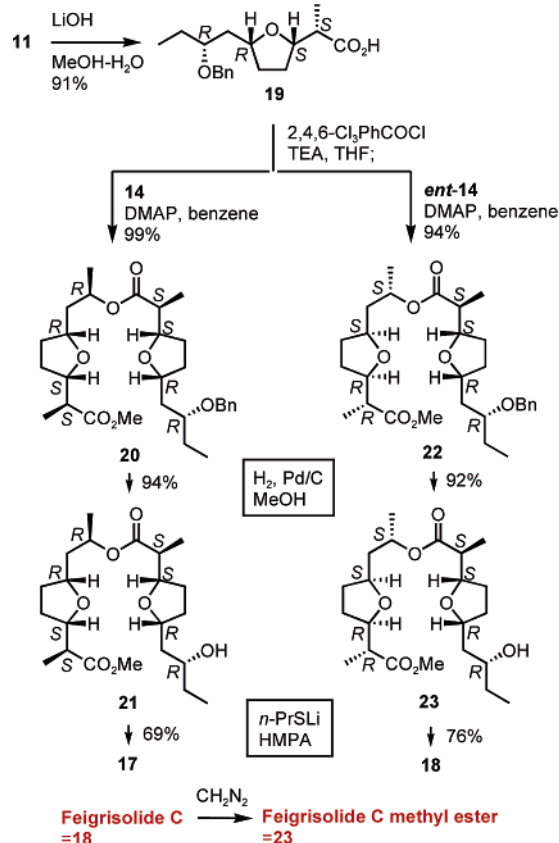
(NMR Analysis) A=15 or *ent*-15  
B=13 or *ent*-13

candidate **18** via **22** and **23**. Careful comparison studies revealed that **18** exhibited spectroscopic characteristics identical to those of feigrisolide C. The synthetic sample of **18** had the same optical rotation value ( $[\alpha]^{25}_D +16.2$  (*c* 0.2, MeOH)) as that reported for feigrisolide C ( $[\alpha]^{20}_D +17.2$  (*c* 0.4, MeOH)), and the true structure of feigrisolide C was finally identified as **18** (Scheme 3). A small sample of feigrisolide C was converted into the methyl ester via reaction with diazomethane, and it exhibited spectroscopic properties identical to those of **23**,<sup>9</sup> confirming the conclusion.

A literature search revealed that bonactin<sup>10</sup> (isolated from a *Streptomyces* sp. BD21-2 cultured from a shallow

(9) In <sup>1</sup>H NMR spectra, the features around  $\delta$  1.7 were diagnostic in differentiating between **17** and **18** and also their methyl esters **21** and **23**.

SCHEME 3. Synthesis of the Possible Structures of Feigrisolide C



water sediment sample,  $[\alpha]^{25}_D 0$  (*c* 0.84, CH<sub>2</sub>Cl<sub>2</sub>)), has the same gross structure as that of feigrisolide C, but it was claimed to consist of a racemic mixture of ( $\pm$ )-nonactinic acid and ( $\pm$ )-homononactinic acid.<sup>11</sup> A nonactoyl homononactinic acid of unknown stereochemistry was also recently claimed to be isolated from a strain of *Streptomyces globisporus*.<sup>12</sup> Determination of the exact structure of each of these compounds is quite difficult, and the present studies suggest expedient ways of solving the problems.

Experimental Section

**Ester 22.** To a mixture of carboxylic acid **19** (63 mg, 0.21 mmol) and TEA (0.065 mL, 0.48 mmol) in THF (2.5 mL) was added 2,4,6-trichlorobenzoyl chloride (0.05 mL, 0.35 mmol). The reaction mixture was stirred for 2 h at room temperature. The white precipitate that formed was removed by filtration under N<sub>2</sub> via cannula transfer to a glass pipet equipped with a septum and a plug of glass wool. The THF filtrate was evaporated by a stream of N<sub>2</sub>. The residue was diluted with benzene (5 mL), and DMAP (75 mg, 0.63 mmol) was added. To this mixture was added alcohol *ent*-**14** (65 mg, 0.30 mmol) in benzene (2.5 mL). The reaction mixture was stirred for 2 h at room temperature, and the reaction was quenched with saturated NH<sub>4</sub>Cl solution (8 mL). The reaction mixture was extracted with DCM (20 mL  $\times$  3), and the organic extracts were dried over MgSO<sub>4</sub>, filtered, and concentrated. Purification by flash column chromatography (hexane–EtOAc, 2:1) gave ester **22** (98 mg (oil), 94%). *R*<sub>f</sub> 0.50

(10) Schumacher, R. W.; Talmage, S. C.; Miller, S. A.; Sarris, K. E.; Davidson, B. S.; Goldberg, A. *J. Nat. Prod.* **2003**, *66*, 1291–1293.

(11) This claim is probably erroneous if bonactin is a single compound.

(12) Rezanaka, T.; Spížek, J.; Příkrylová, V.; Prell, A.; Dembitsky, V. M. *Tetrahedron* **2004**, *60*, 4781–4787.

(hexane–EtOAc, 2:1).  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.36–7.27 (m, 5H), 5.04–4.93 (m, 1H), 4.54 and 4.48 (AB q, 2H,  $J = 11.3$  Hz), 4.07–3.94 (m, 3H), 3.92–3.83 (m, 1H), 3.67 (s, 3H), 3.58–3.51 (m, 1H), 2.55–2.43 (m, 2H), 2.02–1.92 (m, 4H), 1.77–1.72 (m, 2H), 1.69–1.42 (m, 8H), 1.23 (d, 3H  $J = 6.3$  Hz), 1.09 (d, 6H,  $J = 7.0$  Hz), 0.92 (t, 3H,  $J = 7.4$  Hz);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  175.2, 174.2, 139.0, 128.2, 127.7, 127.3, 80.3, 80.1, 78.1, 76.6, 76.5, 71.5, 69.2, 51.5, 45.7, 45.3, 42.5, 40.8, 31.5, 31.3, 28.4, 28.3, 27.1, 20.5, 13.2, 13.1, 9.2. IR (neat):  $\nu_{\text{max}} = 3450, 2972, 2877, 1738, 1497, 1456, 1377, 1259, 1196, 1167, 1061\text{ cm}^{-1}$ . MS  $m/z$  (FAB, relative intensity): 505 ( $\text{M}^+ + 1$ , 49), 397 (10), 307 (24), 199 (100), 154 (64), 136 (46), 91 (79). HRMS (FAB) calcd for  $\text{C}_{25}\text{H}_{45}\text{O}_7$  ( $\text{M}^+ + 1$ ) 505.3165, found 505.3169.  $[\alpha]^{25}_{\text{D}} -15.0$  (c 2.75,  $\text{CHCl}_3$ ).

**Alcohol 23.** Ester **22** (38 mg, 0.073 mmol) was dissolved in MeOH (3.2 mL), and palladium on activated carbon (10% w/w, 16 mg) was added. The mixture was stirred under hydrogen atmosphere for 30 min and then filtered through a filter paper. After solvent evaporation, flash column chromatography (hexane–EtOAc, 2:1) yielded alcohol **23** (28 mg (oil), 92%).  $R_f$  0.41 (hexane–EtOAc, 1:3).  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  5.05–4.94 (m, 1H), 4.16–4.12 (m, 1H), 4.02–3.95 (m, 2H), 3.93–3.84 (m, 1H), 3.78–3.67 (m, 1H), 3.69 (s, 3H), 2.79 (d, 1H  $J = 4.8$  Hz), 2.56–2.44 (m, 2H), 2.04–1.94 (m, 4H), 1.80–1.43 (m, 10H), 1.24 (d, 3H  $J = 6.2$  Hz), 1.10 (d, 6H,  $J = 7.0$  Hz), 0.93 (t, 3H,  $J = 7.4$  Hz);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  175.3, 174.2, 80.8, 80.4, 76.6, 76.4, 70.3, 69.4, 51.6, 45.4, 45.3, 42.4, 40.7, 31.3, 30.6, 30.1, 28.6, 28.5, 20.5, 13.3, 13.2, 10.1. IR (neat):  $\nu_{\text{max}} = 2974, 1732, 1462, 1379, 1263, 1198, 1063\text{ cm}^{-1}$ . MS  $m/z$  (FAB, relative intensity): 415 ( $\text{M}^+ + 1$ , 64), 391 (27), 307 (19), 282 (68), 199 (54), 154 (100), 136 (71), 107 (25). HRMS (FAB) calcd for  $\text{C}_{22}\text{H}_{39}\text{O}_7$  ( $\text{M}^+ + 1$ ) 415.2696, found 415.2706.  $[\alpha]^{25}_{\text{D}} +6.0$  (c 0.53,  $\text{CHCl}_3$ ).

**(2'S,3'S,6'R,8'R)-Homomonactoyl (2R,3R,6S,8S)-Nonactic Acid (18).** To a solution of alcohol **23** (28 mg, 0.067 mmol) in HMPA (0.27 mL) was added lithium *n*-propyl mercaptide (0.48 M in HMPA, 0.027 mL). The reaction mixture was stirred for 2 h at room temperature. The reaction was quenched by addition

of saturated  $\text{NaHCO}_3$  (0.15 mL) solution and water (0.11 mL). The reaction mixture was extracted with  $\text{CHCl}_3$  (0.11 mL  $\times$  6). The aqueous phase was acidified to pH 1 with 2 N HCl and extracted with  $\text{CHCl}_3$  (0.22 mL  $\times$  8). The organic extracts were dried over  $\text{MgSO}_4$ , filtered, and concentrated. Purification by flash column chromatography ( $\text{CHCl}_3$ –MeOH, 9:1) gave (2'S,3'S,6'R,8'R)-homomonactoyl (2R,3R,6S,8S)-nonactic acid (**18**, 21 mg (oil), 76%).  $R_f$  0.40 ( $\text{CHCl}_3$ –MeOH, 9:1).  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  5.09–5.03 (m, 1H), 4.17–4.11 (m, 1H), 4.05–3.91 (m, 3H), 3.80–3.72 (m, 1H), 2.52–2.46 (m, 2H), 2.04–1.97 (m, 4H), 1.83–1.46 (m, 11H), 1.25 (d, 3H  $J = 6.3$  Hz), 1.17 (d, 3H,  $J = 7.0$  Hz), 1.11 (d, 3H,  $J = 7.0$  Hz), 0.93 (t, 3H,  $J = 7.4$  Hz);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  176.8, 174.3, 81.1, 80.5, 77.2, 76.7, 70.4, 68.9, 45.4, 44.9, 42.2, 40.3, 31.0, 30.4, 29.8, 29.2, 28.8, 20.3, 13.7, 13.5, 10.1. IR (neat):  $\nu_{\text{max}} = 3444, 2970, 2933, 2873, 1730, 1460, 1379, 1263, 1198, 1061\text{ cm}^{-1}$ . MS  $m/z$  (FAB, relative intensity): 401 ( $\text{M}^+ + 1$ , 47), 307 (20), 289 (10), 199 (25), 154 (100), 136 (76), 107 (33). HRMS (FAB) calcd for  $\text{C}_{21}\text{H}_{37}\text{O}_7$  ( $\text{M}^+ + 1$ ) 401.2539, found 401.2543.  $[\alpha]^{25}_{\text{D}} +16.2$  (c 0.24,  $\text{CHCl}_3$ ).

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**Supporting Information Available:** Experimental procedures,  $^1\text{H}$  NMR spectra of **A**, **B**, **3**, **13**, **15**, **21**, **23**, feigrisolide C, and feigrisolide C methyl ester, and  $^{13}\text{C}$  NMR spectra of the intermediates. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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